

RAT POISONS NOT ONLY KILL WILDLIFE: THEY CAN ALSO WEAKEN AND SICKEN THEM. Known "sublethal" impacts include:

- Hemorrhaging beneath the skin and extensive bruising. Internal hemorrhaging in bones, body wall, heart, and elsewhere in the body. Possible heart failure.¹
- Hemorrhaging of the heart, liver, kidney, lung, intestines, and muscles.²
- Anticoagulants associated with inflammatory response and immune suppression in bobcats.³
- Anticoagulants associated with multiple system effects in bobcats.⁴
- Multiple AR exposure events associated with notoedric mange.⁵
- Barn owl clutch size, brood size, fledging success, and nest box occupancy lower in fields treated with bromadiolone and chlorophacinone.⁶
- Increased vulnerability to other causes of death such as vehicular collisions and predation.⁷
- Coyotes exposed to multiple FGARs and with high FGAR residues tended to be in poorer body condition.⁸
- Chronic anemia, making animals more susceptible to diseases, including mange, and other stressors.
- Reproductive impacts. Female sheep exposed to anticoagulants had more aborted or stillborn lambs (up to 50%); male sheep had lower sperm motility.¹⁰
- Decreased food intake¹¹ and decreased body weight.¹²
- Neonatal transfer to young kits. Decreased resilience to environmental stressors.¹³ Fetuses more susceptible to brodifacoum toxicity than adults.¹⁴

- Increased parasite and pathogen burdens.¹⁵
- Shorter wings, tails, bones, bills, and birth defects.¹⁶
- Rodents poisoned by anticoagulants are more likely to be eaten by predators.¹⁷
- Raptors may preferentially prey upon sickened rodents: The energetically beneficial behavior of favoring substandard prey may increase raptor encounters with rodenticide exposed animals if prey vulnerability has resulted from poisoning.¹⁸
- Exposure to brodifacoum may have prolonged effects that increase toxicity of subsequent AR exposure.¹⁹
- Bromadiolone and chlorophacinone residues from secondary poisoning can be transferred to the eggs of *T. alba.* ²⁰
- Increased stress and reduced body condition.²¹
- Chlorophacinone can affect ability of hawks to thermoregulate.²²
- Bromadiolone exposure reduced body condition in common kestrel nestlings.²³
- Clutch size, hatching success, and fledging success in barn owls is lower in oil palm plantations treated with warfarin and brodifacoum, relative to untreated plantations.²⁴
- Male barn owls engage in greater exploratory flight and energy expenditure to secure enough prey to meet their needs, when nesting in areas treated with brodifacoum and warfarin, relative to untreated areas.²⁵
- Red foxes that die from or with infectious diseases have a greater disposition to second generation anticoagulant rodenticide exposure.²⁶

¹ Mendenhall and Pank. 1980. Secondary Poisoning of Owls by Anticoagulant Rodenticides. Wildlife Society Bulletin 8:311-315

² Rattner et al. 2011. Acute Toxicity, Histopathology, and Coagulopathy in American Kestrels (Falco sparverius) Following Administration of the Rodenticide Diphacinone. Environmental Toxicology and

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³ Serieys, et al. 2018. Urbanization and anticoagulant poisons promote immune dysfunction in bobcats. Proc Biol Sci. 2018 Jan 31; 285(1871): 20172533

⁴ Fraser, et al. Genome-wide expression reveals multiple systemic effects associated with detection of anticoagulant poisons in bobcats (Lynx rufus) Mol Ecol. 2018;00:1–18.

⁵ Serieys, et al. Anticoagulant rodenticides in urban bobcats: exposure, risk factors and potential effects based on a 16-year study. Ecotoxicology (2015) 24:844–862

⁶ Salim, et al. 2014. Sub-lethal effects of the anticoagulant rodenticides bromadiolone and chlorophacinone on breeding performances of the barn owl (Tyto alba) in oil palm plantations. Slovak Raptor Journal 8(2): 113-122

⁷ Fournier-Chambrillon, et al. 2004. Evidence of Secondary Poisoning of Free-Ranging Riparian Mustelids by Anticoagulant Rodenticides in France: Implications for Conservation of European Mink (Mustela letreola). Journal of Wildlife Diseases 40(4):688-695

⁸ McKenzie, et al. 2022. Exposure of Urban Coyotes to Anticoagulant Rodenticides in Southern California: Sub-lethal Effects and Environmental Correlates. Proceedings of the Vertebrate Pest Conference, 30(30)

⁹ Riley, et al. 2007. Anticoagulant Exposure and Notoedric Manage in Bobcats and Mountain Lions in Urban Southern California. Journal of Wildlife Management 71(6).

¹⁰ Robinson, et al. 2005. Effect of the anticoagulant, pindone, on the breeding performance and survival of merino sheep, Ovis aries. Comparative Biochemistry and Physiology, Part B 140:465-473.

¹¹ Oliver and Wheeler 1978. The toxicity of the anticoagulant pindone to the European rabbit, Oryctogulas cuniculus and the sheep, Ovis aries. Australian Wildlife Research 5:135-142.

¹² Rattner et al. 2011. Acute Toxicity, Histopathology, and Coagulopathy in American Kestrels (Falco sparverius) Following Administration of the Rodenticide Diphacinone. Environmental Toxicology and Chemistry 30(5): 1213-1222

¹² Litten, et al. 2002. Behavior, coagupathy and pathology of brushtail possums (Trichosurus vulpecula) poisoned with brodifacoum. Wildlife Research 29:259-267.

¹³ Gabriel, et al. Anticoagulant Rodenticides on our Public and Community Lands: Spatial Distribution of Exposures and Poisoning of a Rare Forest Carnivore. PLoS ONE 7(7):e40163.

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¹⁵ Lemus, et al. 2011. Side effects of rodent control on non-target species: Rodenticides increase parasite and pathogen burden in great bustards. Science of the Total Environment 409 (2011) 4729-4734

¹⁶ Naim, et al. 2010. Growth Performance of Nesting Barn Owls, Tyto Alba javanica in Rat Baiting Area in Malaysia. J. Agric. Biol. Sci. 5(6):1-13.

¹⁷ Cox and Smith. 1992. Proc. 15th Vertebrate Pest Conf. UC Davis. Rodenticide Ecotoxicology: Pre-Lethal Effects of Anticoagulants on Rat Behavior

¹⁸ Vyas, et al. 2017. Influence of Poisoned Prey on Foraging Behavior of Ferruginous Hawks. Am. Midl. Nat. (2017) 177:75–83

¹⁹ Rattner, et al. 2019. Brodifacoum Toxicity in American Kestrels (*Falco sparverius*) with Evidence of Increased Hazard Upon Subsequent Anticoagulant Rodenticide Exposure. Environmental Toxicology and Chemistry 2020;39(2):468-481.

 $^{^{20}}$ Salim, et al. 2015. The Effects of Rodenticide Residues Deposited in Eggs of *Tyto alba* to Eggshell Thickness. Sains Malaysiana 44(4)(2015): 559–564

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